

Available online at http://www.journalijdr.com



International Journal of Development Research Vol. 06, Issue, 07, pp.8371-8374, July, 2016

Full Length Research Article

CULTURAL, BIOCHEMICAL AND HAEMOLYTIC PROPERTIES OF *STAPHYLOCOCCUS AUREUS* ISOLATED FROM CASES OF SUBCLINICAL MASTITIS IN CATTLE

*Taruna Bhati, Prerna Nathawat, Sandeep Kumar Sharma, Priyanka Rathore and Anil Kumar Kataria,

Department of Vety., Microbiology and Biotechnology, College of Veterinary and Animal Sciences, Rajasthan University of Veterinary and Animal Science, Bikaner, India

ARTICLE INFO	ABSTRACT
Article History:	In the present study 38 Staphylococcus aureus isolates from subclinical mastitis cases in Holstein-

Received 25th April, 2016 Received in revised form 24th May, 2016 Accepted 16th June, 2016 Published online 31st July, 2016

Key Words: Subclinical mastitis, Staphylococcus aureus, Coagulase, Hemolysis, Cattle. In the present study 38 *Staphylococcus aureus* isolates from subclinical mastitis cases in Holstein-Friesian (H-F) crossbred and Rathi (a native breed) cattle were characterized on the basis of their cultural, biochemical and haemolytic properties after confirmation by 23S r RNA based genotyping. The overall incidence of subclinical mastitis was 44.70 % with higher incidence (51.61 %) in H-F crossbred cattle than in Rathi cattle (40.74 %). Among the cultural and biochemical properties, 35 out of 38 isolates produced golden yellow colonies and three isolates produced white colonies. In the sugar fermentation reactions using 11 different sugars, variations were observed in the results. Thirty five isolates produced coagulase and the overall strongest coagulation reaction was recorded with plasma from human followed by horse, cattle, rabbit, sheep, camel, goat, dog, buffalo and chicken. Thirty two isolates were haemolytic while six were non-haemolytic. All the isolates produced α toxin of high titres (1:2560 and 1:5120) and β -toxin was produced by 34 isolates with lower titres (1:5 to 1: 160). None of the isolates produced delta toxin. In conclusion, incidence of subclinical mastitis was more in H-F crossbred cattle than in Rathi cattle. Not all the isolates were pigmented, coagulase producer and hemolytic.

Copyright©2016, *Taruna Bhati et al.* This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

Staphylococcus aureus is a well recognized pathogen leading to both clinical and subclinical bovine mastitis (Suleiman *et al.*, 2012). It has been implicated in more than 80% of subclinical bovine mastitis resulting in economic losses of about 300 \$ per year per animal (Karahan *et. al.*, 2011). Further milk from such animals with subclinical mastitis may legally be sold for human consumption which may lead to many different subsequent infections or food poisoning as such (Oliveira *et al.*, 2011). *Staphylococcus aureus* can be easily isolated from subclinical mastitis cases using nutrient agar and Mannitol Salt Agar as selective media in the laboratory. Hemolysins produced by *S. aureus* have been considered true virulent factors in causation of the disease (Dinges *et al.*, 2000; Ariyanti *et al.*, 2011) and typing and

*Corresponding author: Taruna Bhati,

Department of Vety. Microbiology and Biotechnology, College of Veterinary and Animal Sciences, Rajasthan University of Veterinary and Animal Science, Bikaner, India. titration of these hemolysins may well be an indicator of pathogenicity factor because of its hemolytic, dermonecrotic and neurotoxic effects (Dinges *et al.*, 2000). It produces coagulase, an extracellular enzyme that binds to protein to form a complex with thrombin-like activity which converts fibrinogen to fibrin (McDevitt *et al.*, 1992). The production of coagulase by *S. aureus* has been related to pathogenicity of this organism and has also been used as an important criterion for the phenotypic identification of the organism by many workers. Hence the present study was undertaken for isolation, identification and phenotypic characterisation of genotypically confirmed *S. aureus* isolates from subclinical mastitis in cattle in terms of cultural and biochemical properties, coagulase production, haemolytic properties and toxin assays.

International Journal of

DEVELOPMENT RESEARCH

MATERIALS AND METHODS

Sampling

Eighty five milk samples were collected during early morning hours in sterilized test tubes from Holstein–Friesian (H-F)

crossbred (n=31) and Rathi cattle (n=54) from different locations in Bikaner (Rajasthan, India). The samples were immediately taken to the laboratory for further processing on ice

Somatic cell counting (SCC)

A 0.1ml amount from each properly shaken milk samples was withdrawn with Pasteur pipette and spread evenly on a glass slide to count the somatic cell count as per the method of Prescott and Breed, (1910).

Ribotyping of organisms

The isolates were genotypically confirmed by 23S rRNA species specific PCR using forward primer-1 (5'-ACGGAGTTACAAAGGACGAC-3') and reverse primer-2 (5'-AGCTCAGCCTTAACGAGTAC-3') (Straub *et al.*, 1999).

Identification of S. aureus

All the milk samples which showed SCC corresponding to subclinical mastitis were processed for isolation of *S. aureus* by conventional methods. Milk samples were streaked onto 5% sheep Blood Agar (BA), nutrient agar and DNase agar plates and incubated aerobically at 37°C for 24-48 h. The tests of sucrose, D-mannose, D-manitol, maltose, sucrose, lactose, raffinose, dextrose, arabinose, dulcitol and inositol fermentation were carried out. The tube coagulase test was carried out for of free enzyme using plasmas from different animal species (*viz.* cattle, buffalo, sheep, goat, horse, dog, rabbit, chicken, camel) and human. All these tests were performed as described by Quinn *et al.* (1994).

Haemolytic properties and Haemolysin assays

The hemolytic activity was evaluated by inoculating S. aureus isolates in the form of streaks on the surface of triplicate plates of blood agar base supplemented with 5% sheep, bovine and horse blood for alpha, beta and delta hemolysin assays, respectively (Quinn et al., 1994) and incubated at 37°C for 24 and 48 h. The criteria for hemolysin identification were: complete lytic zone (transparent) with blurred edges for α hemolysin on ovine and incomplete (non-transparent) lytic zone, which became complete with sharp edges after overnight incubation at 4°C on bovine blood agar, for beta hemolysin. The delta-hemolysin production was determined as complete hemolytic zones on horse blood agar (Quinn et al., 1994; da Silva *et al.*, 2005). Qualitative and quantitative assays for α , β and δ haemolysins were done using rabbit, cattle and horse erythrocytes respectively (Sanjiv and Kataria, 2007). The preparation of erythrocytes, typing and titration of haemolysins were done as per method described by (Sanjiv and Kataria, 2007).

RESULTS

Out of the 85 milk samples, 38 milk samples showed SCC in the range of 200×10^3 to 500×10^3 cells/ml corresponding to subclinical cases of mastitis as per the IDF 2005 criterion. All 38 isolates were also confirmed to be *S. aureus* by species specific PCR targeting 23S rRNA and revealed an amplicon of

1,250 bp. In the present investigation, the incidence of subclinical mastitis associated with S. aureus was 44.7%. The recovery of S. aureus was 51.61% from H-F crossbred and 40.74% from Rathi subclinical mastitic milk samples. Following inoculation and culture on nutrient agar, 35 isolates produced golden yellow pigmentation, the intensity of which increased with passage of time whereas three (7.8%) isolates produced white colonies belonging to both the breeds of cattle (one to H-F crossbred and two to Rathi). All the 38 isolates were subjected to aerobic cultivation on mannitol salt agar the color of the medium changed to yellow from original pink by all the isolates but a variation in reaction time was observed for different isolates. In the sugar fermentation reactions using 11 different sugars only two sugars (mannitol and dextrose) were fermented by all the isolates obtained from crossbreds whereas six sugars (mannitol, sucrose, dextrose, fructose, lactose and mannose) were fermented by all the isolates from Rathi cattle. Of the total 38 S. aureus isolates, 100% fermented mannitol and dextrose, 97.4% fermented fructose, sucrose and lactose, 94.7% fermented mannose, 92.1% fermented maltose, 13.1% fermented arabinose and 2.6% fermented raffinose sugar. Dulcitol and inositol were not fermented by any of the isolates from both the cattle breeds. Out of 38 isolates, 31(81.6%) S. aureus isolates showed a positive DNase test. In the tube coagulase test, 35 isolates showed positive coagulase test and three of the isolates showed negative reaction with plasma from all the species used. The overall coagulase reaction in the present study in descending order of superiority was human > horse > cattle > rabbit > sheep > camel > goat > dog > buffalo > chicken. In the present study, 32 (84.2%) isolates were hemolytic and six (15.8%) did not produce haemolysis on blood agar. Out of 32 isolates, six (18.75%) produced complete haemolysis, 17 (53.1%) produced partial haemolysis and nine (28.1 %) of the isolates showed both types of haemolysis on sheep blood agar. Only one isolate showed turning of partial hemolysis to complete hemolysis (hot-cold lysis) whereas other 16 isolates did not show hotcold lysis. In the hemolysin assay, all the 38 isolates haemolysed rabbit erythrocytes indicating presence of alphatoxin whereas beta-toxin was found to be produced by only 34 (89.5%) isolates and none of the isolates produced delta toxin. The titres of alpha toxin produced by most of the S. aureus isolates from crossbred and Rathi cattle were 1:2560. The titres of beta toxin were much below (1:5 to1:160) than that of alpha toxin.

DISCUSSION

The criterion of isolation of organisms along with SCC more than two lacs cells per ml of milk from normal udders has been followed in the present investigation to define subclinical mastitis (SCM). The SCC has been detected to be the most reliable test and closest to the bacteriological results for SCM in dairy cows by. Most of the workers (Sharma *et al.*, 2010; Elango *et al.*, 2010; Moges *et al.*, 2011; Mosaferi *et al.*, 2012) have detected a positive correlation between SCC and bacterial pathogen. The isolation of predominantly *S. aureus* from SCM cases in present study shows concordance between our observation and those from other workers *viz.* Abdel-Rady and Sayed, (2009) Moges *et al.* (2011) and Suleiman *et al.* (2012). The higher incidence of SCM in H-F crossbred than in the Rathi cattle supports the earlier observations of Abdel-Rady and Sayed (2009) and Moges et al. (2011) who also reported higher prevalence of SCM in crossbred breeds than in the indigenous zebu cattle. Difference in colony pigmentation of S. aureus isolates which were recovered from cattle, human and other domestic animals was reported by many workers [Quinn et al. (1994), Adesiyun et al. (1999) Salasia et al. (2004)] as seen in our study. In the present study of the three isolates producing white colonies on nutrient agar, two were coagulase positive in tube test and possessed *coa* gene (as studied by genotyping) whereas the third one did not produce coagulase and was coa gene deficient. Our study revealed that coagulase production was independent of colony pigmentation. The observation of white colonies is in complete agreement to the observations of Sanjiv et al. (2008) who reported one isolate and white colonies and absence of coa gene.

The fermentation reactions in the present study for sucrose, mannose, fructose and lactose were also almost similar to those observed by Khichar (2011). When the results for two cattle breeds were compared it was concluded that isolates of Rathi origin showed fermentation of more sugars included in the study than the isolates from crossbred cattle. Our observations in regards to fermentation of glucose by all the isolates and of inositol by none of the isolate is in complete agreement to the findings of Morandi et al. (2009) who carried out biochemical profiling or metabolic fingerprinting of the S. aureus isolates from dairy products using biologue GP-2 microplate. DNase activity is important to distinguish between pathogenic staphylococci and nonpathogenic resident flora. DNase is as important as coagulase for pathogenesis (Pfaller & Herwaldt, 1988). In the study of Devriese and Oeding (1975), it was found that there was a strong association of DNase and coagulase production for S. aureus. Both of these tests were positive 96% of the time. Contrarily, our study revealed that that there is no correlation between DNase activity and coagulase production with pathogenicity in S. aureus isolates from both the breeds. In the tube coagulase test using plasma from nine species and human, our results were in conformity to those of Adesiyun and Shehu (1985) who also recorded strongest reaction with human and rabbit plasma followed by pig, donkey, chicken, cattle, duck and goat. They also reported spontaneous clotting of horse plasma but it was not recorded in the present study. Our results also supported observations of Kateete et al. (2010) who found that human plasma was more sensitive (91%) than sheep plasma (81%) for the tube coagulase test. The variations in coagulase reaction is probably due to affinity for different plasma samples (Wilson and Miles, The prevalence of β -hemolysis in bovine S. 1975). aureus strains in the present study is in full agreement with Aarestrup et al. (1999); Larsen et al. (2002); Morandi et al., 2009. Our study on pattern of hemolysis of 38 S. aureus isolates on sheep blood agar is in contrast to the observations of Singh (2006) ; Sanjiv and Kataria (2007); Upadhyay and Kataria, (2010); Khichar (2011) who did not record ahemolytic S. aureus from milk of cattle and goat. Similar to our study, Yadav et al., (2015) reported ahaemolytic S. aureus isolates from milk of cattle and buffalo with clinical mastitis. Production of α -toxin by all the isolates in the present study is similar to the findings of Upadhyay and Kataria (2010); Khichar (2011); Yadavet al., (2015) who also reported production of α -toxin by all the isolates from bovine and goat

mastitic milk. The non-production of δ -toxin is in contrast to observations of Upadhyay and Kataria (2010) and Yadav et al. (2015) who demonstrated production of δ -toxin by some of the isolates of S. aureus from mastitic milk samples. The present findings of α -toxin titres of 1:2560 of S. aureus from Rathi and H-F crossbred cattle are in agreement to those of Khichar (2011), Upadhyay and Kataria (2010) who also recorded similar α -toxin titres. The highest titres of 1:5120 for α -toxin are similar to the findings of Sanjiv and Kataria (2007) and Yadav et al., (2015). The highest titres for β -toxin in S. aureus isolates from both the breeds was same being 1:160 which is in complete agreement to the findings of Upadhyay and Kataria (2010) who also reported similar highest titres for β -toxin in isolates from cattle and goat mastitis. In the present study on 38 S. aureus isolates from subclinical mastitic cases, a lot of variations were observed in the biochemical properties since the biochemical reactions of staphylococci have been shown to vary within the same gland over time (Maisi and Riipinen, 1991). In conclusion, incidence of subclinical mastitis was more in H-F crossbred cattle than in Rathi cattle and not all the isolates were pigmented, coagulase producer and hemolytic. Hence different biotypes were observed on the basis of cultural, biochemical and hemolytic properties.

Acknowledgements

The authors are thankful to Dean, College of Veterinary and animal Science, Bikaner for providing facilities to accomplish this work.

Conflict of interest

The Authors declare that there is no conflict of interest.

REFERENCES

- Aarestrup, F. M.; Larsen, H. D., Eriksen, N. H. R., Elsberg, C. S. and Jensen, N. E. 1999. Frequency of α and β -haemolysin in *Staphylococcus aureus* of bovine and human origin. A comparison between pheno- and genotype and variation in phenotypic expression. *Acta Pathologica, Microbiologica et Immunologica Scandinavica*, 107 (4): 425–430.
- Abdel-Rady, A. and Sayed, M. 2009. Epidemiological studies on subclinical mastitis in dairy cows in Assiut governorate. *Vet. World.*, 2, 373-380
- Adesiyun, A. A.; Webb, L. A. and Romain, H. 1999. Phenotypic Charactersitics of *Staphylococcus aureus* strains isolated from milk and dairymen on dairy farms in Trinidad. *Isr J Vet Med.*, 54, 11-17.
- Adesiyun, A. A. and Shehu, L. M. 1985. Detection of staphylocoagulase using plasmas from various animals. Vet Microbiol., 10, 387-392.
- Ariyanti, D.; Salasia, S.I.O. and Tato, S. 2011. Characterization of haemolysin of *Staphylococcus aureus* isolated from food of animal origin. *Indones. J. Biotechnol.*, 16(1): 32-37.
- Da Silva, E. R. and da Silva, N. 2005. Coagulase gene typing of *Staphylococcus aureus* isolated from cows with mastitis in southeastern Brazil. *Can J Vet Res.*, 69, 260-264.

- Devriese, L. A. and Oeding, P. 1975. Coagulase and heatresistant nuclease producing Staphylococcus epidermidis strains from animals. *J Appl Bacteriol.*, 39(2):197–207
- Dinges, M.M.; Orwin, P.M. and Schlievert. P.M. 2000. Exotoxins of *Staphylococcus aureus*. *Clin Microbiol Rev.*,13, 16-34.
- Elango, A., Doraisamy, K. A., Rajarajan, G. and Kumaresan, G. 2010. Bacteriology of subclinical mastitis and antibiogram of isolates recovered from cross breed cows. *Indian J Anim Res.*, 44(4): 280-284.
- International Dairy Federation, (IDF). 2005. Diagnostic potential of California Mastitis Test to Detect Subclinical Mastitis 26. IDF, Maastricht, Netherlands. P:15-19.
- Karahan, M., Aciki, M.N. and Cetinkaya, B. 2011. Investigation of virulence genes by PCR in *Stapylococcus aureus* isolates originated from subclinical bovine mastitis in Turkey. *Pak. Vet. J.*, 31(3): 249-253.
- Kateete, D. P.; Kimani, C. N.; Katabazi, F. A.; Okeng, A.; Okee, M. S.; Nanteza, A.; Joloba, M. L. and Najjuka, F.C. 2010. Identification of *Staphylococcus aureus*: DNase and Mannitol salt agar improve the efficiency of the tube coagulase test. *Ann Clin Microbiol Antimicrob.*, 9, 23.
- Khichar, V. 2011. Genotypic characterization of *Staphylococcus aureus* from clinical mastitis in cattle in relation to some virulence factors. M.V.Sc. Thesis submitted to Rajasthan University of Veterinary and Animal Sciences, Bikaner.
- Maisi, P. and L. Riipinen, 1991. Pathogenicity of different species of staphylococci in caprine udder. *British Vet. J.*, 147, 126-132.
- McDevitt, D.; Vaudaux, P. and Foster, T. J. 1992. Genetic evidence that bound coagulase of *Staphylococcus aureus* is not clumping factor. *Infect Immun.*, 60(4): 1514–1523.
- Moges, N.; Asfaw, Y. and Belihu, K. 2011. A cross sectional study on the prevalence of sub-clinical mastitis and associated risk factors in and around Gondar, Northern Ethiopia, *Int J Anim Vet Adv.*, 3(6): 455-459.
- Morandi, S. ; Brasca, M. ; Andrighetto, C. ; Lombardi, A. and Lodi R. 2009. Phenotypic and Genotypic Characterization of *Staphylococcus aureus* Strains from Italian Dairy Products. *Int J Microbiol.*, 7 pages. Doi :10.1155 /2009 /501362.
- Mosaferi, S.; Jalili, T.; Ostadi, Z.; Khakpour, M. and Bodaghi, H. 2012. Sensitivity of *Staphylococcus aureus* isolated from subclinical bovine mastitis to co-amoxiclav in Tabriz dairy herd in 2010. *Res J Biol Sci.*, 7(4), 165-169.
- Oliveira, L.; Ana, C..; Rodrigues, A. C.; Hulland, C. and Ruegg, P. L. 2011. Enterotoxin production, enterotoxin gene distribution, and genetic diversity of *Staphylococcus aureus* recovered from milk of cows with subclinical mastitis. *Afr J Vet Res.*, 72(10): 1361-1368.
- Pfaller, M. A., and Herwaldt, L. A. 1988. Laboratory, clinical and epidemiological aspects of coagulase negative staphylococci. *Clin Microbiol.*, 37, 201–205.

- Prescott, S.C. and R.S. Breed. 1910. The determination of the number of body cells in milk by a direct method. *J. Infect. Dis.*, **7**:632.
- Quinn, P.J., Carter, M.E., Markey, B.K. and Carter, G.R. 1994. Clinical Veterinary Microbiology. Wolfe Publishing, Mosby-Year Book Europe Ltd., England.
- Salasia, S.I., Khusnan, Z., Lammler, C. and Zschock, M. 2004 Comparative studies on phenol - and genotypic properties of *Staphylococcus aureus* isolated from bovine sub-clinical mastitis in central Java in Indonesia and Hesse in Germany. J. Vet. Sci., 5(2): 103-109.
- Sanjiv, K. and Kataria, A.K. 2007. Typing and titration of haemolysins produced by *Staphylococcus aureus* of cattle mastitis origin. J. Anim. Health., 46(1): 51-55.
- Sanjiv, K.; Kataria, A. K.; Sharma, R. and Singh, G. 2008. Epidemiological typing of *Staphylococcus aureus* by DNA restriction fragment length polymorphism of *coa* gene. *Vet Arhiv.*, 78 (1), 31 -38.
- Sharma, N., Pandey, V. and Sudhan, N.A. 2010. Comparison of some indirect screening tests for detection of subclinical mastitis in dairy cows. *Bulg. J. Vet. Med.*, 13: 98-103.
- Singh, J. 2006. Bacterial determinants of sub-clinical cattle mastitis with special reference to *Staphylococcus aureus*. M.V.Sc. thesis submitted to the Rajasthan Agric. Univ., Bikaner, Rajasthan.
- Straub J A, Hertel C and Hammes W P. 1999. A 23S rRNA target polymerase chain reaction based system for detection of *Staphylococcus aureus* in meat starter cultures and dairy products. *J Food Protect.*, 62, 1150-56
- Suleiman, A.B., Kwaga, J.K.P., Umoh, V.J., Okolocha, E.C., Muhammed, M., Lammler, C., Shaibu, S.J., Akinden, O. and Weiss, R. 2012. Macro-restriction analysis of *Staphlyococcus aureus* isolated from subclinical bovine mastitis in Nigeria. *Afr. J. Microbiol. Res.*, 6(33): 6270-6274.
- Upadhyay, A. and Kataria, A.K. 2010. Haemolytic properties and titration of haemolysins of *Staphylococcus aureus* of milk origin from cattle and goat with clinical mastitis. Ind. *J Vet. Res.*, 19(2): 60-65.
- Wilson, G. S. and Miles, A. 1975. Topley and Wilson's principles of bacteriology, virology and immunity, E. Arnold, London. pp. 781-83.
- Yadav, R. Sharma, S. K., Yadav, J. Bhati, T. and Kataria, A. K. 2015. Phenotypic and Genotypic Haemolysin Properties of *Staphylococcus aureus* Obtained from Milk of Cattle and Buffalo with Clinical Mastitis. *J Pure Appl Microbiol.*, Vol. 9(1), p: 349-355.

8374