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REVIEW ON POSITIVE ROLE OF REACTIVE OXYGEN SPECIES (ROS) IN SEED GERMINATION

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ABSTRACT

In many seeds, the process of germination features testa rupture and endosperm rupture are two separate events. Endosperm rupture requires cell wall weakening in the endosperm layer. Puncture force measurements are a useful tool for quantifying this endosperm weakening. Various mechanisms have been proposed to promote endosperm weakening. Endosperm weakening requires cell wall weakening. This process involves cleavage of cell wall polymers or loosening of bonds between the polymers. Several molecular mechanisms have been proposed for endosperm weakening. Most prominent among them is, cleavage of cell wall polymers in the endosperm by reactive oxygen species, or more specifically, by apoplastic hydroxyl radicals ('OH) formed when superoxide (O_2^{-*}) and hydrogen peroxide (H_2O_2) undergo a Fenton reaction in the presence of peroxidases. Regulation of seed germination is quite complex and is further complicated by interaction of hormones like Gibberellin (GA), Abscisic Acid (ABA) and ethylene. Moreover, the involvement of reactive oxygen species (ROS) in hormone signaling for such regulation is still less understood. Positive interactive role of ROS with GA, ABA and ethylene was observed in seed germination of Vignaradiata. ROS production is essential for lignification and cross-linking of cell wall polymers in vascular tissue. Oxidation of the germination inhibitor(s) present in the pericarp by H2O2 promotes seed germination. Antioxidants which are derivatives of well-known germination inhibitors suppressed seed germination in a dose-dependent manner. To initiate seed germination, the germination inhibitor(s) should be decomposed by an oxidant such as H₂O₂

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INTRODUCTION

ROS are reactive molecules and Free Radicals derived from molecular oxygen. They are chemically reactive molecules containing oxygen (H_2O_2 , O_2^- , OH). These ROS were initially recognized as toxic by-products of aerobic metabolism but it also involved in "Positive" developmental role and a novel direct action during seed endosperm weakening and germination. Seeds are reported to generate ROS either in dry condition or even when imbibed (Bailly, 2004) and ROS generation has been associated with a positive role in the process of germination (Chaudhuriet al., 2008; Garnczarska and Wojtyla, 2008; Ishibashiet al., 2010). Seed germination, an early developmental event, starts with imbibitional water uptake and culminates into radicle protrusion. Seed germination involves activation of embryonic growth following hydration. Underlying this activation is a metabolic up regulation that includes some subtle intangible regulatory

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components like Reactive Oxygen Species (ROS) which are connected with messengers like hormones through complex signalling chains (Kar, 2007). The highly reactive free radicals, along with hydrogen peroxide (H₂O₂), are collectively termed active/reactive oxygen species (AOS/ROS). While the intracellular glassy state must curtail molecular and AOS mobility, and thus interaction, during the process of desiccation, intracellular structures are highly vulnerable as conditions for radical generation are enhanced (Vertucci and Farrant, 1995; Pammenter and Berjak, 1999; Walters et al., 2005). For an endospermic seed to complete germination, the growth potential of the radicle must be high enough to overcome the tissue resistance of the endosperm (Bewley, 1997). Factors that influence germination, e.g. plant hormones, can thus influence the resistance of the endosperm tissue by promoting or inhibiting endosperm weakening. Increased cellular levels of Reactive Oxygen Species (ROS) are known to occur during seed development and germination (Bailly, 2004). Reactive Oxygen Species (ROS) were initially recognized as toxic by-products of aerobic metabolism, removed by means of antioxidants and antioxidative enzymes.

So, most studies on ROS and antioxidants have been focused on negative of role in seeds like seed deterioration followed by ageing. In recent years, it has become apparent that ROS play an important positive role like endosperm weakening and seed germination.

Mechanism on Formation of Reactive Oxygen Species

NADH oxidases located in the plasma membrane catalyse the formation of apoplastic O2-anions. O2- is dismutated by the antioxidant enzyme Superoxide Dismutase (SOD), leading to the formation of H₂O₂and molecular oxygen. Thus O₂-•and H₂O₂are both present in the apoplast. In the presence of bivalent cations (e.g. Fe2+or Cu2+), •OH can be formed from H₂O₂in the so-called Fenton reaction, and regeneration of these catalytic bivalent cations can be achieved by the oxidation of O_2 .-• (Vianello and Macri, 1991). The generation of •OH in the cell wall by a Fenton-type reaction can also take place in the presence of peroxidases, which are abundant in the plant cell wall (Chen and Schopfer, 1999), or in the presence of ascorbate and bivalent cations (Fry, 1998). •OH areable to cleave hemicelluloses and have been shown to cause invitrocleavage of cell wall polysaccharides (Fry, 1998; Schweikert et al., 2002).



Fig. 1.Qualitative model for the generation of apoplastic hydroxyl radicals. NADH oxidases located in the plasma membrane catalyse the formation of apoplastic O_2 .-•anions. O_2 .-• is dismutated to H_2O_2 and molecular oxygen by superoxide dismutase (SOD). A Fenton-type reaction can take place in the presence of peroxidases, leading to the formation of •OH. (From Schopfer *et al.*, 2001.)

H₂O₂reacts with O-• in the presence of cell wall peroxidases, leading to the formation of •OH (Chen and Schopfer, 1999). These •OH then act on the cell wall by causing polysaccharide cleavage resulting in endosperm weakening. Peroxidases are generally present in cell walls of growing tissues in great abundance, OH can potentially be generated there whenever H₂O₂ and O₂ are available. Peroxidase activity increases in the micropylar endosperm of tomato seeds prior to endosperm rupture (Morohashi, 2002). Ascorbate peroxidase scavenging H₂O₂ is localized at the site of H₂O₂ generation in the plant cell (Asada, 1999), and is likely to be micro compartmentalized by organizing scavenging complex of such enzymes as superoxide dismutase and monodehydroascorbatereductase (Ogawa et al., 1995, Asada et al., 1996, Asada, 1999). During the biosynthesis of lignin, H₂O₂ is produced at the site of lignification (Ogawa et al., 1996, Ogawa et al., 1997). Thereby, H₂O₂ generated is scavenged or utilized at its generation site in plants. If H_2O_2 is generated at an unexpected site or reaches a level that exceeds the scavenging capacity, it causes oxidative damage to plants. H_2O_2 impairs the photosynthetic activity of isolated chloroplasts when applied exogenously even at $1\mu M$ which is a level having little effect on the activity when generated in chloroplasts (Asada and Takahashi, 1987).

Role of Ros in Cell wall Loosening

Endosperm weakening followed by seed germination requires cell wall weakening. This process involves cleavage of cell wall polymers or loosening of bonds between the polymers. Several molecular mechanisms have been proposed for endosperm weakening. Digestion of cell wall polysaccharides by hydrolytic enzymes is one mechanism. The main focus so far has been on β -1,4-mannanase (Bewley, 1997) and β -1,3glucanase (Leubner-Metzger, 2003). Reactive Oxygen Species (ROS) produced in the cell wall play a role in endosperm weakening, which takes place before endosperm rupture. Peroxidase activity increases in the micropylar endosperm of tomato seeds prior to endosperm rupture (i.e. during endosperm weakening) (Morohashi, 2002). The cleavage of cell wall polymers by •OH not only takes place in the endosperm, but also plays a role in radicle elongation. The cell walls have to be loosened in order to allow cell elongation, caused by water uptake, which takes place when the water potential in the embryo is lower than that of the surrounding medium (Muller et al., 2007).

In tobacco seeds, b-1,3-glucanase is induced in the micropylar part of the endosperm prior to radicle protrusion suggested that the enzyme contributes to the weakening of the cell walls of the micropylar endosperm by helping to hydrolyse cell wall b-1,3-glucans and thereby contributes to facilitating penetration of the radicle (Vogeli-Lange et al., 1994 and Leubner-Metzger et al., 1995). Non-enzymatic cleavage of wall polymers by the hydroxyl radical (OH). This extremely reactive, short-lived intermediate of O₂ reduction is principally capable of splitting covalent bonds in all kinds of organic molecules in a diffusion-limited reaction, i.e. within a few nanometers of its site of formation. As their range of action is limited by diffusion, they must be produced directly in the cell wall in order to cleave cell wall polymers (Halliwell and Gutteridge, 1989; Schopfer, 2001). Cell-wall polysaccharides such as pectin and xyloglucan can be broken down in vitro by OH generated in a Fenton-type reaction, e.g.by the reduction of O₂ with ascorbate in the presence of Cu ions.

OH produced in the apoplastic space of plant tissues, could act as a site-specific oxidant targeted to play a useful physiological role in cell-wall loosening processes underlying cell expansion, fruit ripening and organ abscission. (Fry, 1998). Radicle penetration through the micropylar portion of the endosperm may accompany the breakdown of the cell wall, that is, wounding (Wu *et al.*,2001). Many studies have shown that peroxidase is induced by wounding in plants (Desbiez and Boyer, 1981; Espelie *et al.*, 1986; Kawaoka *et al.*,, 1994; Kato *et al.*, 2000). On the other hand, peroxidase has been reported to be involved in generation of H_2O_2 from NADH (Maeder *et al.*, 1980). Inhibition of catalase in lettuce seeds led to higher concentrations of H_2O_2 in the seeds and to faster germination (Hendricks and Taylorson, 1975).

Interaction of Hormones with Ros

Regulation of seed germination is quite complex and is further complicated by interaction of hormones like Gibberellin (GA), Abscisic Acid (ABA) and ethylene. Moreover, the involvement of Reactive Oxygen Species (ROS) in hormone signaling for such regulation is still less understood. Among phytohormones, GA and ABA are well known for their antagonistic action for seed dormancy and germination. They play a contrasting role GA breaks dormancy and promotes germination while ABA maintains dormancy and inhibits germination (Bentsink and Kooenneef, 2002). Recently, Reactive Oxygen Species (ROS) have been demonstrated to play roles in growth and development either directly by participating in the process of cellular growth or differentiation or indirectly by signaling for induction of processes or reactions related to growth and differentiation (Bailly, 2004).

ROS may act downstream of plant hormones (Kwak, 2003). H2O2 rescued germination almost fully from the inhibition imposed by ABA and ABA might have affected ROS metabolism either directly or by interfering with ethylene action on ROS metabolism (Chaudhuri et al., 2013). Sarath et al. (2007) also demonstrated a reversal of ABA-induced inhibition of germination by H2O2 and they explained this as due to interference of H2O2 in ABA signalling. On the other hand, action of GA on seed germination of V. radiatais in line with H₂O₂, as H2O2 can partially overcome the inhibition by paclobutrazole on germination. Roles of ROS in GAsignalling in the aleurone layer and Programmed Cell Death (PCD) in Hordeumvulgare has already been established, where GA initiates cell death of aleurone cells, whereas ABA inhibits cell death (Ishibashi et al., 2013). GA may have some other way of signalling in use for germination as H₂O₂ could not fully rescue when GA synthesis was blocked by paclobutrazole (Chaudhuri et al., 2013). Liu et al. (2010) demonstrated that H₂O₂upregulates ABA catabolism through NO signalling while promotes GA synthesis thus favouring germination. On the other hand, Bahin et al. (2011) proposed that H₂O₂alleviates dormancy by activating GA signalling and synthesis, not by repressing ABA signalling.

Germination Promoting Mechanism of Ros

It is known that H_2O_2 promotes germination in various species. H₂O₂ promoted seed germination in a dose-dependent manner as did respiratory inhibitors, indicating that H2O2 itself possibly promotes seed germination rather than O_2 . H_2O_2 is a toxic molecule due to its highly oxidative reactivity and long life. In the presence of catalytic metal ions such as iron and copper, it produces hydroxyl radical (·OH) which strongly oxidizes cell components such as membrane lipids and enzymes.H₂O₂ can be used in high concentrations to promote germination of seeds with hard seed coats by scarification, it also has a germination-promoting effect in lower concentrations (Naredo et al., 1998; Ogawa and Iwabuchi, Exogenously applied H₂O₂ ameliorates 2001). seed germination in many plants (Fontaine et al., 1994, Chien and Lin, 1994). This has been explained by the fact that the scavenging activity for H₂O₂ is high enough, resulting in the production of O₂ for mitochondrial respiration. However, an early step of seed germination (dormancy breakage) dispenses

with mitochondrial O₂ respiration and requires the activation of the oxidative pentose phosphate pathway. Thioredoxin reduction by NADPH produced via the oxidative pentose phosphate pathway allows the mobilization of storage proteins of cereals, leading to germination (Fontaine et al., 1994). The pericarp and seedcoat often contain phenolic compounds and alkaloids which inhibit seed germination (Bhattacharyya et al., 1999, Tao andButa 1986). Oxidation of germination inhibitor(s) is likely to exclude a reaction that is mediated by peroxidases because strong inhibitors of peroxidases that have micromolar Ki value promoted seed germination. It is likely that H₂O₂oxidatively denatures germination inhibitor(s) such as ferulic and coumaric acids. The germination frequency of Z. elegans seeds was enhanced by H2O2. The concentration of H₂O₂ giving the maximal promotional effect varied depending on the time after imbibition of the seeds. The concentrations of H_2O_2 that gave half the maximal germination frequency achieved 24 h and 48 h after imbibition were obtained between 2 and 5 mM, whereas it was between 5 and 20 mMH₂O₂ at 36 h and 48 h after imbibition.

These suggest that H₂O₂ influences more than two steps in the process of germination and/or is continuously consumed at a certain rate to promote seed germination. The disproportionation of H₂O₂ resulting in an increased O2 level is considered to enhance the oxidative respiration, which can be the reason for seed germination promotion (Ogawa, 2001). Imbibition of dry seeds is associated with a rapid increase in oxygen uptake and mitochondrial respiration supporting ATP synthesis (Bewley and Black, 1985). It is estimated that up to 2% of mitochondrial O₂ consumption in seeds is involved in the generation of H₂O₂ (Cakmak et al., 1993). High concentrations of O₂ generally enhance the germination of plant seeds, and such promotion has also been considered to be due to the enhancement of mitochondrial oxidative respiration. The K_mvalue for mitochondrial Cyt*c*oxidase which participates in the ATP production metabolism accompanied with O2 consumption is estimated to be approximately 140 nM O₂, whereas that for alternative oxidase whose reaction is not accompanied with ATP production is approximately 1.7 µM O₂ (Millar et al., 1994).

Germination inhibitor(s) may block other processes involved in germination aside from mitochondrial respiration. Activation of the oxidative pentose phosphate pathway generally leads to seed germination, and has been proposed to be promoted by H₂O₂ (Fontaine et al., 1994). The oxidative pentose phosphate pathway provides NADPH which is used for the reduction of redox-regulating proteins such as thioredoxin. Such proteins regulate mobilization of storage proteins and the modulation of activities of enzymes and transcriptional factors by their reduction of disulfide bond in the target molecules (Kobrehel et al., 1992). So, the activation of the pentose phosphate pathway leads to germination. Antioxidant germination inhibitors such as phenolic compounds may block activation of the pathway. Catalase and ascorbate peroxidase activities increase and dehydroascorbatereductase activity decreases with germination of wheat seeds. So, there is low scavenging activity of H₂O₂ in seeds at the initial stage of germination and the temporal oxidized state of the seed embryo that is induced by H₂O₂ might initiate germination and that antioxidant germination inhibitor(s) might prevent the induction of the oxidized state in seeds (Cakmaket al., 1993).

REFERENCES

- Asada K., 1999. The water-water cycle in chloroplasts: Scavenging of active oxygens and dissipation of excess photons, *Annu. Rev. Plant Physiol., Plant Mol. Biol.*, 50: 601–639.
- Asada K., and Takahashi M., 1987, Production and scavenging of active oxygen in photosynthesis. In Photoinhibition, Edited by Kyle, D.J., Osmond, C.B. and Arntzen, C.J. pp. 227–287, Elsevier Science Publishers, Amsterdam.
- Asada K., Miyake C., Ogawa K., and Hossain M.A., 1996, Microcompartmentation of ascorbate peroxidase and regeneration of ascorbate form ascorbate radical: Its dual role in chloroplasts. In Plant Peroxidases: Biochemistry and Physiology. Edited by Obinger, C., Burner, U., Ebermann, R., Penel, C. and Greppin, H. pp. 163–167. University of Geneva.
- Bahin E., Bailly C., Sotta B., Kranner I., Corbineau F., 2011, Crosstalk between reactive oxygen species and hormonal signalling pathways regulates grain dormancy in barley, *Plant Cell Environ*, 34: 980-993.
- Bailly C., 2004, Active oxygen species and antioxidants in seed biology, *Seed Sci Res.*, 14: 93-107.
- Bentsink L., Kooenneef M., 2002, Seed dormancy and germination, In: Somerville CR, Meyerowitz EM (eds) The Arabidopsis Book, *American Society of Plant Biologists, Rockville*, MD, doi/10.1199/tab.0009.
- Bewley D.J., and Black M., 1985, Cellular events during germination and seedling growth, In Seeds, Physiology of Development and Germination, pp. 135–173, Plenum Press, London.
- Bewley J.D., 1997, Breaking down the walls a role for endoβ-mannanase in release from seed dormancy? *Trends in Plant Science*2, 464–469.
- Cakmak I., Strbac D., and Marschner H., 1993, Activities of hydrogen peroxide-scavenging enzymes in germinating wheat seeds, J. Exp. Bot., 44: 127–132
- Chaudhuri A., Kar R.K., 2008, Inhibition of seed germination by propyl gallate, a free radical scavenger and recovery of germination by hydrogen peroxide and ethylene in Vigna radiate, *World J Agri Sci.*, 4: 914-921.
- Chaudhuri A., Singh K.L., and Kar R.K., 2013, Interaction of Hormones with Reactive Oxygen Species in Regulating Seed Germination of Vignaradiata(L.) Wilczek. J Plant Biochem Physiol., 1:1.
- Chen S., and Schopfer P., 1999, Hydroxyl-radical production in physiological reactions: a novel function of peroxidase, *European Journal of Biochemistry*, 260, 726–735.
- Desbiez M.O., Boyer N., 1981, Hypocotyl growth and peroxidases of Bidenspilosus. *Effect ofcotyledonaryprickings and lithium pretreatment, Plant Physiology*, 68, 41-43.
- Espelie K.E., Franceschi V.R., Kolattukudy P.E., 1986. immunocytochemical localization and time-course appearance of an anionic peroxidase associated with suberization in wound- healing potato tuber tissue, *Plant Physiology*, 81, 487- 492.
- Fontaine O., Huault C., Pavis N., and Billard, J.P., 1994, Dormancy breakage of Hordeumvulgareseeds: Effects of hydrogen peroxide and scarification on glutathione level and glutathione reductase activity, *Plant Physiol. Biochem.*, 32: 677–683.

- Fry S., 1998, Oxidative scission of plant cell wall polysaccharides by ascorbate-induced hydroxyl radicals, *Biochemical Journal*, 332, 507–515.
- Garnczarska M., Wojtyla L., 2008, Differential response of antioxidative enzymes in embryonic axes and cotyledons of germinating lupine seeds, *ActaPhysiol Plant.*, 30: 427-432.
- Halliwell B., and Gutteridge J.M.C., 1989, Free Radicals in Biology and Medicine, 2ndedn. Oxford, UK: Clarendon Press.
- Hendricks S., and Taylorson R., 1975, Breaking of seed dormancy by catalase inhibition. Proceedings of the National Academy of Sciences of the United States of America, 72, 306–309.
- Ishibashi Y., Tawaratsumida T., Zheng S-H., Yuasa T., Iwaya-Inoue M., 2010, NADPH oxidase act as key enzyme on germination and seedling growth in barley (Hordeumvulgare L.)., *Plant Physiol.*, 13: 45-52.
- Ishibashi Y., Koda Y., Zheng S.H., Yuasa T., Iwaya-Inoue M., 2013, Regulation of soybean seed germination through ethylene production in response to reactive oxygen species. *Ann Bot.*, 111: 95-102.
- Kar R.K., 2007, Physiology and metabolic regulation of seed germination. In: Trivedi PC (ed) Plant Physiology: Current Trends, Pointer Publishers, Jaipur, India, 290-304.
- Kato M., Hayakawa Y., Hyodo H., Ikoma Y., Yano M., 2000, Wound-induced ethylene synthesis and expression and formation of 1-1minocylopropane-1-carboxylate (ACC) synthase, ACC oxidase, phenylalanine ammonia-lyase, and peroxidase in wounded mesocarp tissue of Cucurbita maxima. *Plant and Cell physiology*, 41, 440-447.
- Kawaoka A., Kawamoto T., Ohta H., Sekine M., Takano M., Shinmyo A., 1994, Wound-induced expression of horseradish peroxidase, *Plant Cell Reports*, 13, 149-154.
- Kobrehel K., Wong J.H., Balogh A., Kiss F., Yee B.C., and Buchanan B.B., 1992, Specific reduction of wheat storage proteins by thioredoxinh., *Plant Physiol.*, 99: 919–924.
- Kwak J.M., Mori I.C., Pei Z.M., Leonhardt N., Torres M.A., 2003, NADPH oxidase AtrohD and AtrohF genes function in ROS-dependent ABA signaling in Arabidopsis, *EMBO J*, 22: 2623-2633.
- Leubner-Metzger G., Frundt C., Vogeli-Lange R., MeinsJr F., 1995, class I b-1-3-glucanase in the endosperm of tobacco during germination. *Plant Physiology* 109,751-759.
- Leubner-Metzger G., 2003, Functions and regulation of β-1,3glucanase during seed germination, dormancy release and after-ripening. *Seed Science Research*, 13, 17–34.
- Liu Y., Ye N., Liu R., Chen M., Zhang J., 2010, H2O2 mediates the regulation of ABA catabolism and GA biosynthesis in Arabidopsis seed dormancy and germination, *J Exp Bot.*, 61: 2979-2990.
- Maeder M., Ungemach J., Schloss P., 1980. The role of peroxidase isozyme groups of Nicotianatabacum in hydrogen peroxide formation, *Planta.*, 147, 467-470
- Millar A.H., Bergersen F.J., and Day D.A., 1994, Oxygen affinity of terminal oxidases in soybean mitochondria, *Plant Physiol., Biochem.*,32: 847–852.
- Morohashi Y., 2002, Peroxidase activity develops in the micropylar endosperm of tomato seeds prior to radicle protrusion, *Journal of Experimental Botany.*, 53, 1643–1650.

- Muller, K., Hess B., Leubner-metzger G., 2007. Role of reactive oxygen species in endosperm weakening, Seed biology, *development and ecology.*, pp 287-295.
- Ogawa K., and Iwabuchi M., 2001. A mechanism for promoting the germination of Zinnia elegans seeds by hydrogen peroxide, *Plant Cell Physiology*, 42, 286–291.
- Ogawa K., Kanematsu S., and Asada K., 1997. Generation of superoxide anion and localization of CuZn-SOD in the vascular tissue of spinach hypocotyls: Their association in lignification, *Plant Cell Physiol.*, 38: 1118–1126.
- Ogawa K., Kanematsu S., Takabe K., and Asada K., 1995. Attachment of CuZn-superoxide dismutase to thylakoid membranes at the site of superoxide generation (PS I) in spinach chloroplasts: Detection by immuno-gold labeling after rapid freezing and substitution method, *Plant Cell Physiol.*, 36: 565–573.
- Pammenter N.W., and Berjak P., 1999. A review of recalcitrant seed physiology in relation to desiccationtolerance mechanisms, *Seed Science Research*, 9, 13–37.
- Sarath G., Hou G., Baird L.M., Mitchell R.B., 2007, Reactive oxygen species, ABA and nitric oxide interactions on the germination of warm-season C4-grasses, *Planta*, 226: 697-708.
- Schopfer P., 2001. Hydroxyl radical-induced cell-wall loosening in vitro and in vivo: implications for the control of elongation growth, *The Plant Journal*, 28, 679–688.

- Schopfer P., Plachy C., and Frahry G., 2001. Release of reactive oxygen intermediates (superoxide radicals, hydrogen peroxide and hydroxyl radicals) and peroxidase in germinating radish seeds controlled by light, gibberellin, and abscisic acid, *Plant Physiology*, 125, 1591–1602.
- Schweikert C., Liszkay A., and Schopfer P., 2002. Polysaccharide degradation by Fenton reaction- or peroxidase generated hydroxyl radicals in isolated plant cell walls, *Phytochemistry*, 61, 31–35.
- Vertucci C.W., and Farrant, J.M., 1995. Acquisition and loss of desiccation tolerance, In: Kigel, J. and Galili, G. (eds) Seed Development and Germination. Marcel Dekker, New York, pp. 237–271.
- Vianello A., and Macri F., 1991. Generation of superoxide anion and hydrogen peroxide at the surface of plant cells, *Journal of Bioenergetics and Bio membranes*, 23, 409– 423.
- Vogeli-Lange R., Frundt C., Hart C.M., Beffa R., Nage F., MeinsJrF., 1994. Evidence for a role of b 1,3-glucanase in dicot seed germination, *The Plant Journal*, 5, 273-278.
- Walters C., Hill L.M., and Wheeler L.M., 2005. Dying while dry: kinetics and mechanisms of deterioration in desiccated organisms, Integrative and Comparative Biology, 45, 751–758.
- Wu C.T., Leubner-Metzger G., MeinsJr F., Bradford K.J., 2001. Class I b-1,3-glucanase and chitinase are expressed in the micropylar endosperm of tomato seeds prior to radicle emergence, *Plant Physiology*, 126, 1299-1313.
