

ISSN: 2230-9926

Available online at http://www.journalijdr.com



International Journal of Development Research Vol. 07, Issue, 10, pp.16401-16406, October, 2017

# **ORIGINAL RESEARCH ARTICLE**



Open Access

# SALT STRESS-INDUCED PROLINE ACCUMULATION, CHANGES IN IONIC ADJUSTMENT (NA<sup>+</sup>/K<sup>+</sup>) AND CATALASE IN THE TOMATO (*SOLANUM LYCOPERSICUM*) LEAF

# \*Dr. Asha Sharma, Pooja and Govinda

Department of Botany, M.D. University, Rohtak-124001, Haryana, India

#### ARTICLE INFO

Article History: Received 08<sup>th</sup> July, 2017 Received in revised form 28<sup>th</sup> August, 2017 Accepted 14<sup>th</sup> September, 2017 Published online 30<sup>th</sup> October, 2017

Keywords:

Adverse, Ionic, Oxidative, Reactive, Salinity.

#### \*Corresponding author

#### ABSTRACT

Soil salinity is a major limitation to plant in many areas of the world. Present study has explored the effect of salinity on Proline content, mineral composition and antioxidant enzyme of tomato. Tomato plants were exposed to 0, 60, 90,120,150 mM NaCl.The Proline accumulation was studied by reading the absorption of chromophore at 520nm using spectrophotometer. The content of proline increase in tomato as the level of NaCl increase. Using atomic absorption spectrophotometer ion uptake for Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>+</sup> and Mg<sup>2+</sup> were determined. The Na content significantly increase but the content of K<sup>+</sup>, Ca<sup>+</sup>, K<sup>+</sup> and Mg<sup>2+</sup> decrease significantly decrease as the salinity increase. The changes in the activity of antioxidant enzymes such as super oxide dismutase catalase (CAT: EC 1.11.1.6), in leaves of tomato cultivar (cv.) differing in salt tolerance were investigated. The activity of catalase increase as the level of salinity increase.

**Copyright** ©2017, Dr. Asha Sharma, Pooja and Govinda. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

**Citation: Dr. Asha Sharma, Pooja and Govinda, 2017.** "Salt stress-induced proline accumulation, changes in ionic adjustment  $(na^+/k^+)$  and catalase in the tomato (*Solanum lycopersicum*) leaf", *International Journal of Development Research*, 7, (10), 16401-16406.

# **INTRODUCTION**

Vegetables are the good resource for overcoming micronutrient and macronutrient deficiencies and provide small holder farmers with much higher income and more jobs per hectare than staple crops. Tomato is cultivated for its fleshy fruits and it is called as protective food because of its special nutritive value and its wide spread production. Tomato is the richest source of nutrients, dietary fibres antioxidant like lycopene and beta-carotene, the compounds that protect cells from cancer (Hobson, 1993). But excessive salinity reduces the productivity of many agricultural crops including most of the vegetables like tomato. Stress factors based on their origins divided into 2 group's i.e. abiotic and biotic stress factors (Mahaian and Tuteia, 2005). Abiotic stress factors include cold and hot temperatures, drought, salinity, excessive water, radiation, various chemicals, oxidative stress, wind, and lack of soil nutrients. Saline soils are one of the major abiotic stresses that can adversely affect the overall metabolic activities and cause plant death (Roychoudury et al., 2008). About 20% of the world's cultivated land area and 50% of all irrigated land is affected by salinity (Moud and Maghsoudi,

2008). Salt stress affects some major processes such as germination, speed of germination, root/shoot dry weight and  $Na^{+}/K^{+}$  ratio in root and shoot (Parida and Das, 2005). Early flowering reduced dry matter, increased root shoot ratio and leaf size caused by salinity which may be considered as possible ways of decreasing yield in plant under salt stress condition (Mengel et al., 2001). A great contributor to salinity stress is the accumulation of high concentrations of  $Na^+$  in the leaf cell cytoplasm (Jha et al., 2010). Ionic imbalance occurs in cells due to excessive accumulation Na<sup>+</sup> and Cl<sup>-</sup> ions that reduce uptake of  $K^+$ ,  $Ca2^+$ , and Mn (Bayuelo-Jimenez *et* al., 2003). It inhibits the uptake and transport of different ions. Effects of salinity stress on plants have primarily focused on growth, proline accumulation, chlorophyll content, K/Na, Ca/Na ratio, Na<sup>+</sup> and Cl<sup>-</sup>accumulation. The adaptation is generally associated with osmoregulation adjustment by using some osmotic regulators such as potassium, soluble sugar, proline and betaine (Munns, 2005; Hong-Bo et al., 2006). One of the mechanisms is proline accumulation into cell. The role of proline in cell osmotic adjustment, membrane stabilization and detoxification of injurious ions in plants exposed to salt stress is widely reported (Kavi et al., 2005). However the significance of proline accumulation in osmotic adjustment is still debated and varies according to the species (Lutts *et al.*, 1996; Rodriguez *et al.*, 1997). Salinity exerts their effect by causing oxidative damage. This damage is caused by increased production of reactive oxygen species (ROS) (Smimoff, 1995). Excess production of ROS during stress results from impaired electron transport processes in chloroplast and mitochondria as well as from pathways such as photorespiration (Sanchez Rodriguez *et al.*, 2012). In the absence of a protective mechanism in plants ROS can cause serious damage to different aspects of cell structure and function such as initiating lipid per oxidation and damaging DNA, proteins and other small molecules (Arora *et al.*, 2002; Gill and Tuteja, 2010; Ahmad *et al.*, 2011)

Salt tolerance is generally attributed to up-regulated activities of antioxidant enzymes. In tomato salt-tolerance is attributed to the increased activities of superoxide dismutase (SOD), catalase (CAT), ascorbate peroxidase (APX) (Mittova et al., 2004). Earlier it has been reported that the increased  $Na^+$  and decreased K<sup>+</sup> levels leads to Na<sup>+</sup> toxicity (Borsani *et al.*, 2003) and proline accumulation may be due to expression of genes encoding key enzymes of proline synthesis or low activity of the proline oxidizing enzymes (Amini and Ehsanpour, 2005). Enhanced proline accumulation may regulate multiple processes required for survival in salt stress conditions (Maggio et al., 2002). An important aspect of salinity stress is the interaction between Na and K ions (Greenway and Munns, 1980). Sodium provided in low amounts stimulates growth and development of plants and can improve the organoleptic characteristics of the edible parts (Satti et al., 1996). However, high concentrations of Na in the soil inhibit plant growth and reduce commercial yield (Graifenberg et al., 1993, 1996). Potassium regulates the osmotic potential of glycophyte tissues and therefore plays a role in water relations (Lauchli and Pfluger, 1978). Therefore, elevated levels of K could modulate the absorption and the transport of Na and limit the damages attributed to it.

# **MATERIALS AND METHODS**

The plant material which was included in our study was a tomato variety (C-21) purchased from Division of Vegetable Science, IARI, and New Delhi. The seeds were surface sterilized with dilute solution of sodium hypo chlorite (NaOCI) to prevent any fungal contamination and then rinsed three times with distilled water. Then seeds were grown in Petri dishes containing double layered wet filter paper with tap water in order to check the viability of seeds. The seeds were sown in five sets in an earthen pot containing equal quantities (4kg) of loamy sand soil. Salt treatment of NaCl was prepared using sodium chloride salt in concentrations of 60, 90, 120 and 150mM in soil, leaving one set as a control. The samples were taken from two weeks old seedlings for physiological analysis.

## Proline determination in tomato

The proline content in was determined by the method of Bates *et al.*, 1973 using D-Proline as standard.

## Method

Frozen plant material was homogenized in 3% aq. Sulphosalicyclic acid (0.01g/0.5ml) and the residue was removed by centrifugation at 12,000 rpm for 10 mi.Then

added 1 ml of the homogenized tissue in 1 ml of acidninhydrin and 1 ml of glacial acetic acid in a test tube for 1 hr at 100° C and the reaction is terminated in an ice bath. Reaction mixture was extracted with 2ml of toluene mixed vigorously and left at room temperature for 30 min until the separation of two phases .The chromophore containing toluene(1ml,upper phase) was warmed to room temperature an its optical density was measured at 520nm using toluene as a blank. The proline concentration was determined by preparing a standard curve using D-Proline.

## **Mineral and Ionic Composition**

These were obtained from the oven dried material.

#### Sodium and Potassium

200 mg of oven dried and well grinded material was taken in a 50 ml conical flask to which 50 ml of diacid mixture was added ( $H_2SO_4$ &  $HClO_4$ , 4:1). When fumes were heated gentallyon a hot plate till the formation of dense white fumes. When fumes reduced & subsided, heating was increased & digestion was continued for another 25-30 minutes to obtain a colourless digest. The digest was obtained, cooled and diluted to 25 ml distilled water. This acid digest was used further for the estimation of Na & K contents were determined using Flame photometer and expressed as mg g<sup>-3</sup> tissue dry weight. Prior to determination of Na and K contents of tissue digested. It was calibrated using graded concentration of 0-100 ppm solution of Na and K respectively.

Na & K content = O.D.  $\times$  Dilution factor/Wt. of tissue taken = x µg.

## For Quantitative estimation of Catalase in tomato

Tomato sample were taken and crushed in methanol and heated till 80% of methanol evaporates. 10% of extract in 50 ml of buffer (0.067M, pH 7.0 was prepared by mixing 8 ml of A and 42 ml of B in 16:84 ratio, where A contains 1.36g of monobasic sodium sulphate in 50 ml of distilled water and B contains 2.68g of Dibasic sodium phosphate in 50ml of distilled water) followed by addition of  $H_2O_2$  phosphate buffer which is prepared by adding 0.75 ml of 15%  $H_2O_2$  to 50 ml of phosphate buffer (pH 7.0). After 10 min. Read the absorbance was taken at 240 nm (Cakmak and Marschner, 1992).

# **RESULTS AND DISCUSSION**

#### Proline determination in tomato

The proline concentration was increased with increase in NaCl concentration. Also when compared to standard the proline conc. was decreased in sample. The maximum proline concentration was shown in 150mM of NaCl concentration and minimum conc. was shown in control set.

 Table 1a. Analysis of variance table for proline content

Source of Variation	SS	df	MS	F	P-value	F-crit
Between	20888.72	2	10444.36	9.319677	0.003609	3.885294
groups With in	13448.14	12	1120.678			
groups Total	34336.862	14				

Since  $p \le 0.05$ . We can reject the null hypothesis and say that there is an effect of NaCl on proline.

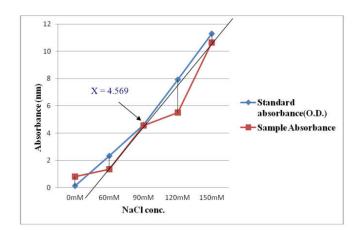


Figure 1. Effect of different concentrations of NaCl on proline content

#### Mineral and ionic composition

#### **Sodium and Potassium**

The sodium content is more than the potassium content with increase in salt concentration. Maximum sodium content and also ratio between two was at 150mM concentration and minimum content was shown in control set or 0mM concentration.

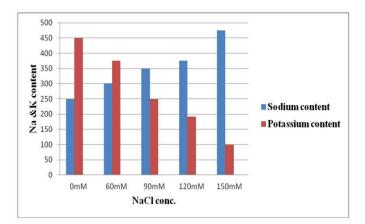


Figure 2a. Effect of different concentration of salt on sodium and potassium content

Table 2a. Analysis of variance table for sodium content

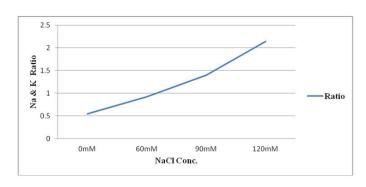
Source of Variation	SS	df	MS	F	P-value	F-crit
Between groups	236922.1	2	118461.1	16.2556	0.000384	3.885294
With in	87448.8	12	7287.4			
groups Total	324370.9	14				

Since  $p \le 0.05$ . We can reject the null hypothesis and say that there is an effect of NaCl on sodium content.

Table 2b. Analysis of variance table for Potassium content.

Source of Variation	SS	df	MS	F	P-value	F-crit
Between	100778.5	2	50389.27	3.630865	0.055468	3.88529
groups With in groups	166536.4	12	13878.03			
Total	267314.9	14				

Since  $p \le 0.05$ . We can reject the null hypothesis and say that there is an effect of NaCl on potassium content



# Figure 2b. Effect of different concentration of salt on sodium and potassium ratio

#### For Quantitative estimation of Catalase in tomato

It was clearly shown from table that the catalase activity was increased with increase in salt conc. The maximum catalase activity was shown at 150mM conc. while minimum was shown ain control set.

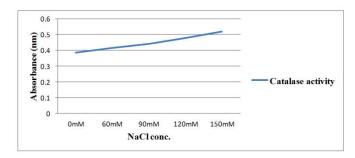


Figure 3. Effect of different concentration of salt on catalase activity

Table 3. Analysis of variance table for catalase activity

Source of Variation	SS	df	MS	F	P-value	F-crit
Between	23261.71	2	11630.86	10.47824	0.00233	3.885294
groups With in	13320.02	12	1110.001			
groups Total	36581.73	14				

Since  $p \le 0.05$ . We can reject the null hypothesis and say that there is an effect of NaCl on catalase activity.

The extent of plant injury by elevated concentration is specific and strongly depends on the environmental conditions and on the availability of salt concentration. In the present study we took the one variety of tomato (C 21). This variety was grown under NaCl salt stress. Different studies on salt stress indicate that salinity have an adverse effect on the growth of tomato plant. In this study we have taken five concentrations (0mM, 60mM, 90mM, 120mM, 150mM) of NaCl were used in order to evaluate its effect on proline, sodium potassium ratio, catalase activity and antioxidant activity. Proline content was also observed after applying NaCl stress. As shown in Table 1 the proline content is increased with NaCl concentration and at 150mM conc. tomato variety show maximum amount of proline content. Significant differences were observed in  $Na^{+}/K^{+}$  ratio among tomato variety under stress treatments. The  $Na^+/K^+$  ratio in taken tomato variety ranged from minimum 0.55 (0mM) to the maximum of 3.80 (150mM) (Fig. 2 a). Salinity stress caused significant increase in  $Na^+/K^+$  ratio. The  $Na^+/K^+$  ratio of the tissues was generally increased with salinity (Amini and Ehsanpour, 2005). Salt stress is characterized by greater Na<sup>+</sup> uptake by the plants and its accumulation in the vacuole (Borsani *et al.*, 2003). The excess of Na<sup>+</sup> could be extremely toxic to the plants. Thus, the salt tolerance of a genotype is determined by its ability to tolerate high Na<sup>+</sup> in shoots without serious effects or keeping high Na<sup>+</sup> in the roots (Chookhampaeng *et al.*, 2007). Increased Na<sup>+</sup> uptake in saline conditions decreases the K<sup>+</sup> uptake due to inhibition power which upset the process of uptake. There is an increase in catalase activity with increase in NaCl conc. as shown in Table – 3 & Fig. – 4. Maximum catalase activity was observed at 150mM NaCl conc. (0.521) which is reduced with decrease in NaCl conc. (0.385) is observed in 0mM conc. of NaCl stress (Control).

#### Acknowledgement

First and foremost, I express my deep sense of gratitude towards Dr. Asha Sharma (Assistant prof.), M.D.University, Rohtak, for her invaluable guidance, cordial advice and constant encouragement throughout this research work. I would like to record my heartfelt gratitude to Prof. Pushpa Dahiya, Head of Botany Department, Faculty of Life Sciences, M.D. University, Rohtak, for rendering her kind co-operation and allowing me to use department facilities to complete my research work.

## REFERENCES

- Ahmad P, Nabi G, Jeleel CA, Umar S. 2011. Free radical production, oxidative damage and antioxidant defense mechanisms in plants under abiotic stress. In: Ahmad P, Umar S (eds) Oxidative stress: role of antioxidants in plants. Studium Press, New Delhi, pp 19–53
- Al-Aghabary K, Zhu Z, Shi Q. 2005. Influence of silicon supply on chlorophyll content, chlorophyll fluorescence, and anti oxidative enzyme activities in tomato plants under salt stress. *J Plant Nutr.*, 12:2101–2115
- Alam, S.M., S.S.M. Naqvi and A.R. Azmi 1989. Effect of salt stress on growth of tomato. *Pak. J. Sci. & Ind. Res.*, 32(2): 110-113
- Amini, F. and A.A. Ehsanpourm, 2005. Soluble proteins, carbohydrates and  $Na^+/K^+$ , changes in two tomato (*Lycopersicon esculentim* Mill.) cultivars under *in vitro* salt stress. *Am. J. Biochem. & Biotechnol.*, 1: 212-216
- Arora A, Sairam RK, Srivastava GC 2002. Oxidative stress and anti oxidative systems in plants. *CurrSci.*, 82:1227– 1238
- Ashraf M 2004. Some important physiological selection criteria for salt tolerance in plants. *Flora*, 199: 361–376
- Ashraf M. 1994. Breeding for salinity tolerance in plants. *Crit Rev Plant Sci.*, 13: 17–42.
- Bates L.S, RP Waldren and D Tear 1973. Rapid determination of free proline for water stress studies. *Plant soil*, 39: 205-207.
- Bayuelo-Jimenez, J.S., D.G. Debouck, and J.P. Lynch, 2003. Growth, gas exchange, water relations and ion composition of *Phaseolus* species grown under saline conditions. *Field Crop Res.*, 80:207–222
- Beck LA. 1984. Case study: San Joaquin Valley Calley Calif. *Agric.*, 41: 16-17
- Bethke, P.C. and M.C. Drew, 1992. Stomatal and nonstomatal components to inhibition of photosynthesis in leaves of *Capsicum annum* during progressive exposure to NaCl salinity. *Plant Physiol.*, 99:219–226

- Bohnert HJ, Jensen RG. 1996. Metabolic engineering for increased salt tolerance: the next step. *Aust J Plant Physiol.*, 23:661–667
- Bolarin, M.c., EG. Cuartero, V. Cruz, and J. Cuartero, 1991. Salinity tolerance in four wild tomato species using vegetative yield-salinity response curves. J. Am. Soc. Hort. Sci., 116:286-290
- Borsani, O., V. Valpuesta and M.A. Botella, 2003. Developing salt tolerant plants in a new century: A molecular biology approach. *Plant Cell Tissue and Organ Culture*, 73: 101-115
- Brand-Willliams W, Cuvelier ME, Berset C. 1995. Use of free radical method to evaluate antioxidant activity. *Lebensm Wiss Technology*, 28:25-30
- Cakmak I, Marschner H. 1992. Magnesium deficiency and high light intensity enhances activities of superoxide dismutase, ascorbate peroxidase and glutathione reductase in bran leaves. *Plant Physiology*, 98: 1222-1227
- Carvajal, M., V. Martinez, C.F. Alcatraz, 1999. Physiological function of water-channels, as affected by salinity in roots of paprika pepper, *Physiol. Plant.*, 105: 95–101
- Chaves, M.M., J. Flexas, and C. Pinheiro, 2009. Photosynthesis under drought and salt stress: Regulation mechanism of whole plant to cell. *Ann. Bot.*, 103:551–568
- Cruz, V., 1. Cuartero, M.c. Bolarin, and M. Romero, Evaluation of characters for ascertaining salt stress' responses in *Lycopersicon* species. 1. Am. Soc. Hort. Sci., 115: 1000-1003 Missing year
- Cuartero J, Fernandez-Munoz R 1990. Tomato and salinity. *Scientia Horticultural*, 78:83–125.
- Cuartero J., Bolarin M.C., Asins M.J. and V. Moreno, 2006. Increasing salt tolerance in the tomato. *J.Exp. Bot.*, 57:1045–1058
- De la Pena R, Hughes J 2007. Improving vegetable productivity in a variable and changing climate. *SAT e Journal* (ejournalicrisatorg) 4 (1):1–22
- Esechie, H.A., A. Al-Saidi and S. Al-Khanjari, 2002. Effect of sodium chloride salinity on seedling emergence in chickpea. J. Agron. And Crop. Sci., 188:155–160
- Evlagon, D., I. Ravina, and P.M. Neumann, 1992. Effects of salinity stress and calcium on hydraulic conductivity and growth in maize seedling roots. J. Plant Nutr., 15(617):795-803
- Fariba A. and A.A. Ehsanpour, 2005. Soluble proteins, proline, carbohydrates and  $Na^+/K^+$  changes in two tomato (*Lycopersicon esculentum* Mill.) cultivars under *in vitro* salt Stress. *Am. J. Biochem. & Biotechnol.*, 1: 212-216.
- Gill S, Tuteja N 2010. Reactive oxygen species and antioxidant machinery in a biotic stress tolerance in crop plants. *Plant Physiol Biochem.*, 48:909–930
- Giovanelli G, Lavelli V, Peri C, Nobili, S. 1999. Variation in ripening. J Sci Food Agric., 79:1583–1588
- Gould WA, 1983. Tomato Production, Processing and Quality Evaluation.2ed.AVI Publishing Company, Inc. Westport, CT. 3-50
- Graifenberg, A., Botrini, L., Giustiniani, L., Lipucci Di Paola, and M. 1996. Salinity affects growth, yield and elemental concentration of fennel. *Hort. Sci.*, 31:1131-1134
- Graifenberg, A., M. Lipucci di Paola, L. Giustiniani, and O. Temperini 1993. Yield and growth of globe artichoke under saline-sodic conditions. *Hort Science* 28:791–793
- Greenway, H. 1973. Salinity, plant growth and metabolism. J. Aust. Inst. Agric. Sci. 393:24-34

- Greenway, H. and Munns, R. 1980. Mechanisms of salt tolerance in non halophytes. Annu. Rev. Plant Physiol. Plant Mol. Biol., 31:149-190
- Hasegawa, P.M., Bressan, R.A., Zhu, J.-K. And Bohnert, H.J 2000b. Plant cellular and molecular responses to high salinity. *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, 51: 463-499).
- Hernàndez JA, Corpas FJ, Gómez M, Del Rio LA, Sevilla F 1993. Salt induced oxidative stress mediated by activated oxygen species in pea leaf mitochondria. *Physiol Plant*, 89:103–110
- Hobson G.E. and D. Grierson, 1993. Tomato. In: Seymour G.B., Taylor J.E., Tucker G. (eds) *Biochemistry of Fruit Ripening*. London, Chapman and Hall: 405-442
- Holden, N 1965. Chlorophylls. In T. W. Goodwin (ed.) Chemistry and biochemistry of plant pigments. Academic Press, N.Y. p. 461-488
- Iyenger, E.R.R. and M.P. Reddy: Photosynthesis in highly salt-tolerant plants, in: M. Pessaraki and M. Dekker. (Eds.) 1996. Handbook of photosynthesis. Marcel Dekker, New York. p: 897–909
- Jain S, Nainawatee HS, Jain RK, Chowdhury JB. 1991. Proline status of genetically stable salt-tolerant *Brassica juncea L*. Somaclones
- Jaleel A, MM Azooz, 2009. Exogenous calcium alters pigment composition, α glutamyl kinase and Proline Oxidase activities in salt stressed *Withaniasomnifera*, *Plant Omics Journal*, 2(2):85-90
- Jha D, Shirley N, Tester M, Roy SJ. 2010. Variation in salinity tolerance and shoot sodium accumulation in Arabidopsis ecotypes linked to differences in the natural expression levels of transporters involved in sodium transport. *Plant, Cell and Environment,* 33: 793–804
- Johnson RW, Dixon MA, and Lee DR. 1992. Water relations of the tomato fruit during growth. *Plant Cell and Environment*, 15: 947-953
- Kabir, M., M.Z. Iqbal, M. Shafiq and Z.R. Farooqi, 2008. Reduction in germination and seedling growth of *Thespesia populnea* L. caused by lead and cadmium treatments. *Pakistan Journal of Botany*, 40(6): 2419-2426
- Kavi KPB, Sangam S, Amrutha RN, Laxmi PS, Naidu KR, Rao KRSS, Rao S, Reddy KJ, Theriappan P, Sreenivasulu N. 2005. Regulation of proline biosynthesis, degradation, uptake and transport in higher plants: Its implications in plant growth and abiotic stress tolerance. *Curr. Sci.*, 88: 424-438
- Khan MA, Irwin A, Allan MS. 2000. The effect of salinity on the growth, water status and ion content of a leaf succulent perennial halophyte, *Suaeda fruticosa L. Arid Environ. J.*, 45: 73-84
- Läuchli, A. and E. Epstein 1990. Plant responses to saline and sodic conditions. In K.K. Tanji (ed). Agricultural salinity assessment and management. ASCE manuals and reports on engineering practice No, 71. pp 113–137
- Lutts S, Kinet JM, Bouharmont J. 1996. Effects of salt stress on growth, mineral nutrition and proline accumulation in relation to osmotic adjustment in rice (*Oryza sativa* L.) cultivars differing in salinity resistance. *Plant Growth Reg.*, 19: 207-218
- Maas, E. V. and S. R. Grattan 1999. Crop yields as affected by salinity. In R. W. Skaggs and J. Van Schilfgaarde (eds) Agricultural Drainage. Agron. Monograph 38. ASA, CSSA, SSA, Madison, WIpp: 55–108
- Maggio, A., S. Miyazaki, P. Veronese, T. Fujita, J.I. Ibeas, B. Damsz, M.L. Narasiman, P.M. Hasegawa, R.J. Joly and

R.A. Bressan 2002. Does proline accumulation plays an active role in stress induced growth reduction? *Plant J.*, 31:699-712

- Mahajan S. and Tuteja N. 2005. Cold, salinity and drought stress: an overview. Arch Biochem Biophys., 444: 139–158
- Marschner, H. 1986. Functions of mineral nutrients: Macronutrients, In: H. Marschner (Ed.). Mineral nutrition in higher plants. Academic, London p: 299–312
- Mittova V, Guy M, Tal M, Volokita M. 2004. Salinity up regulates the antioxidative system in root mitochondria and peroxisomes of the wild salt-tolerant tomato species Lycopersicon pennellii. *J Exp Bot.*, 55:1105–1113
- Mortezai nejad F and P Rezai 2009. Agricultural Research Letters, 1(2): 34-45
- Mugdal V, Madaan N, Mudgal A. 2010. Biochemical mechanism of salt tolerance in plants: A review. *International Journal of Botany*, 6: 136–143
- Munns R, Greenway H, Kirst GO 1983. Halo tolerant eukaryotes. *Physio.J.*, 12: 59-135
- Munns R, R.A. James and A. Lauchli 2006. Approaches to increasing the salt tolerance of wheat and other cereals. *J. Expt. Bot.*, 57:1025–1043
- Munns R. 1993. Physiological processes limiting plant growth in saline soil: some dogmas and hypotheses, *Plant Cell Environ.*, 16: 15–24
- Munns R. 2005. Genes and salt tolerance: bringing them together. *New Phytol.*, 167:645–663
- Munns R. and James RA 2003. Screening methods for salinity tolerance: a case study with tetraploid wheat. *Plant and Soil* 253:201–218
- Munns R. and Tennaat A. 1986. Whole plant responses to salinity. *Aust. J. Plant Physiol.*, 13:143-160.
- Munns R. and Tester M 2008. Mechanisms of salinity tolerance. *Annual Review of Plant Biology*, 59:651–681
- Munns, R 2002. Comparative physiology of salt and water stress. *Plant Cell and Environ.*, 25: 239-250
- Niedziela, Jr., C.E., P.V. Nelson, D.H. Willits, M.M. Peel 1993. Short-tenn salt-shock effects on tomato fruit quality, yield and vegetative prediction of subsequent fruit quality. J. Am. Soc. Hort. Sci., 118:12-16
- Parida AK. and Das AB 2005. Salt tolerance and salinity effects on plants: a review. *Ecotoxical Environ Safety*, 60: 324–349
- Queirosa F, Rodriguesc JA, Almeidaa JM, Almeidad DPF, Fidalgoa F. 2011. Differential responses of the antioxidant defence system and ultrastructure in a salt-adapted potato cell line. *Plant Physiol Biochem.*, 49:1410–1419
- Rahdari P, Tavakoli S, Hosseini SM. 2012. Studying of salinity stress effect on germination, proline, sugar, protein, lipid and chlorophyll content in Purslane (Portulaca oleraceae L.) leaves. *Stress Physio and Bio. J.*, 8(1): 182-193
- Reddy, M.P. and A.B. Vora, 1986. Changes in pigment composition, hill reaction activity and saccharide metabolism in bajra (*Pennisetum typhoides* S&H) leaves under NaCl salinity. *Photosynthetica*, 20:50–55
- Rodriguez HG, Roberts JKM, Jordan WR, Drew MC. 1997. Growth, water relations and accumulation of organic and inorganic solutes in roots of maize seedlings during salt stress. *Plant Physiol.*, 113: 881-893
- Roychoudury A, Basu S Sankar SN and Sengupta DN. 2008. Comparative Physiological and molecular responses of a common aromatic indica rice cultivar to high salinity with non aromatic indica rice cultivar. *Plant cell Rep.*, 27:1395-1410

- Sameni, A.M. and Morshedi, A. 2000. Hydraulic conductivity of calcareous soils as affected by salinity and sodicity. II. Effect of gypsum application and flow rate of leaching solution carbohydrate pol. Soil Sci. *Plant Anal.*, 31: 69-80
- Sánchez-Rodríguez E, Rubio-Wilhelmi M, Blasco B, Leyva R, and Romero L, Ruiz JM. 2012. Antioxidant response resides in the shoot in reciprocal grafts of drought-tolerant and drought-sensitive cultivars in tomato under water stress. *Plant Sci.*, 188–189:89–96
- Sanders, D. 2000. Plant biology: The salty tale of *Arabidopsis*. *Curr. Biol.*, 10: 486-488
- Scholberg, J.M.S., Locascio, S.J. 1999. Growth Response of Snap Bean and Tomato as Affected by Salinity and Irrigation Method, *Hort Science*, 34: 259-264
- Serrano R. and Gaxiola R 1994. Microbial models and salt stress tolerance in plants. *Crit Rev Plant Sci.*, 13: 121–133
- Serrano, R., Culiañz-Macia, A. and Moreno, V. 1999. Genetic engineering of salt and drought tolerance with yeast regulatory genes. *SciHortic.*, 78: 261-269.
- Shalata A. and Tal M 1998. The effects of salt stress on lipid peroxidation and antioxidants in the leaf of the cultivated tomato and its wild salt tolerant relative Lycopersicon pennellii. *Physiol Plant*, 104:169–174

- Shannon, M.C, J.W Gronwald, and M. Tall, 1987. Effects of salinity on growth and accumulation of organic and inorganic ions in cultivated and wild tomatoes. J. Am. Soc. Hort. Sci., 112:416-423
- Shannon, M.C. and C.M. Grieve, 1999. Tolerance of vegetable crops to salinity. *Scientia Horticulturae.*, 78: 5-38.
- Shereen, A., S. Mumtaz, S. Raza, M.A. Khan and S. Soloangi, 2005. Salinity effects on seedling growth and yield component of different rice inbred lines. *Pak. J. Bot.*, 37:131-139
- Smimoff N. 1995. Antioxidant systems and plant response to the environment. In: Smimoff N (ed) Environment and Plant Metabolism. *Bios Scientific Publishers, Oxford, pp*: 217–243.
- Smith, MAL, LA Spomer, RA Shibli, and S.L. Knight, 1992. Effect of NaCl salinity on miniature dwarf tomato 'Micro-Tom': H. Shoot and root growth responses, fruit production and osmotic adjustment. *Plant Nutr.*, 15:2329-2341
- Zhao; K., R Munns; RW King, 1991. Australian Journal of Plant Physiology: 18: 17-24
- Zhu, J.-K., Liu, J. and Xiong, L. 1998. Genetic analysis of salt tolerance in *Arabidopsis*: Evidence for a critical role to potassium nutrition. *Plant Cell.*, 10: 1181-1191.

\*\*\*\*\*\*